Preclinical and clinical evaluation of sulforaphane for chemoprevention in the breast

Brian S.Cornblatt1, Lingxiang Ye2, Albena T.Dinkova-Kostova2, Melanie Erb3, Jed W.Fahey2, Navin K.Singh3, Min-Shue A.Chen4, Tracey Stierer5, Elizabeth Garrett-Mayer6, Pedram Argami2, Nancy E.Davidson4, Paul Talalay2, Thomas W.Kensler1,2,6 and Kala Visvanathan6,7,

1Department of Environmental Health Sciences, 2Department of Pharmacology and Molecular Sciences, 3Department of Surgery, 4Department of Pathology, 5Department of Anesthesiology and Critical Care Medicine, 6Department of Oncology and 7Department of Epidemiology, Johns Hopkins University Bloomberg School of Public Health, 615 North Wolfe Street, Baltimore, MD 21205, USA

To whom correspondence should be addressed. Tel: +1 410 614 1112; Fax: +1 410 614 2632; Email: kvivsana@jhsphs.edu

Consumers of higher levels of Brassica vegetables, particularly those of the genus Brassica (broccoli, Brussels sprouts and cabbage), reduce their susceptibility to cancer at a variety of organ sites. Brassica vegetables contain high concentrations of glucosinolates that can be hydrolyzed by the plant enzyme, myrosinase, or intestinal microflora to isothiocyanates, potent inducers of cytoprotective enzymes and inhibitors of carcinogenesis. Oral administration of either the isothiocyanate, sulforaphane, or its glucosinolate precursor, glucoraphanin, inhibits mammary carcinogenesis in rats treated with 7,12-dimethylbenz[a]anthracene. In this study, we sought to determine whether sulforaphane exerts a direct chemopreventive action on animal and human mammary tissue. The pharmacokinetics and pharmacodynamics of a single 150 µmol oral dose of sulforaphane were evaluated in the rat mammary gland. We detected sulforaphane metabolites at concentrations known to alter gene expression in cell culture. Elevated cytoprotective NAD(P)H:quinone oxidoreductase (NQO1) and heme oxygenase-1 (HO-1) gene transcripts were measured using quantitative real-time polymerase chain reaction. An observed 3-fold increase in NQO1 enzymatic activity, as well as 4-fold elevated immunostaining of HO-1 in rat mammary epithelium, provides strong evidence of a pronounced pharmacodynamic action of sulforaphane. In a subsequent pilot study, eight healthy women undergoing reduction mammoplasty were given a single dose of a broccoli sprout preparation containing 200 µmol of sulforaphane. Following oral dosing, sulforaphane metabolites were readily measurable in human breast tissue enriched for epithelial cells. These findings provide a strong rationale for evaluating the protective effects of a broccoli sprout preparation in clinical trials of women at risk for breast cancer.

Introduction

Cancer chemoprevention entails the use of natural or synthetic agents to retard, block or reverse any of the multiple stages of the carcinogenic process (1–3). It is probable that a combination of approaches will be required to reduce the risk of the major types of human cancers. Current chemopreventive strategies have been directed at individuals in high-risk groups, and may not be suitable for those at moderate or lower risk. This has been observed in the case of breast cancer prevention where the use of tamoxifen, a selective estrogen receptor modulator to prevent hormone receptor-positive breast cancer, has been limited even in women at high risk for breast cancer due to the perception of an unfavorable side-effect profile. As breast cancer remains a global public health challenge affecting one in eight North American and Northern European women, with an increasing incidence in developing countries, the need for non-toxic and inexpensive preventive agents that have broad application is great.

In addition to inherited genetic alterations and estrogen burden, environmental exposures are believed to play a significant role in the development of breast cancer through the cumulative accumulation of mutagenic events over a woman’s lifetime (4). Extent of damage to DNA reflects a molecular equilibrium between activating and detoxifying reactions as well as DNA repair capacity (5,6). This balance can be altered by many factors including drugs and phytochemicals (7). Environmental carcinogens are often first metabolically activated via phase 1 enzymes (principally cytochrome P450) into reactive intermediates (8). Fortunately, at the cellular level, there exist competing phase 2 enzymes (e.g. glutathione-S-transferases and NAD(P)H:quinone oxidoreductase (NQO1)) which facilitate the elimination of the reactive forms of carcinogens through biotransformation reactions including sulfation, acetylation, quinone reduction and glutathione conjugation (9). As a consequence, a promising strategy for cancer chemoprevention involves the use of agents to favorably shift the balance between phase 1 and 2 enzymes (3,5,10).

Such a favorable enzymatic induction strategy, in which cytoprotective enzymes are selectively elevated, has been demonstrated using Brassica vegetables that contain compounds known as glucosinolates (β-thioglucoside N-hydroxysulfates) (11). Glucosinolates are hydrolyzed to isothiocyanates by myrosinase (β-thioglucoside glucohydrolase), an enzyme released in the plant upon damage, such as by chewing (12). Sulforaphane, one such isothiocyanate whose precursor is abundant in 3-day-old broccoli sprouts, triggers the induction of many cytoprotective enzymes including NQO1 and heme oxygenase-1 (HO-1). NQO1 protects cells from oxidative damage by two-electron reduction of quinones, such as those derived from estrogens, thus suppressing oxidative cycling and reactive oxygen species generation (13). HO-1, an important enzyme in heme catalysis, leads to the production of biliverdin, which upon reduction forms the reactive oxygen scavenger bilirubin (14). Much of sulforaphane’s cytoprotective enzyme induction is thought to occur via the actions of the transcription factor nuclear factor erythroid 2-related factor 2 (NRF2) (15). Importantly, when sulforaphane or its precursor, glucoraphanin, was given to female rats treated with the known mammary gland carcinogen, 7,12-dimethylbenz[a]anthracene, the number, size and rate of development of mammary tumors were significantly reduced (16,17). Additional preclinical studies revealed the capacity of sulforaphane to diminish DNA adduct formation in normal mammary epithelial cells, favorably adjust cellular redox states, and inhibit mitosis and proliferation in mammary cancer cell lines (18–22). These laboratory findings are supported by some epidemiological data that suggest a protective association between increased intake of broccoli or Brussels sprouts and breast cancer (23–25).

The identification of biomarkers for efficacy and breast cancer risk is necessary to advance diet-derived chemopreventive agents in both animal and human studies (26). Several clinical studies using 3-day-old broccoli sprouts have already established their safety, described plasma pharmacokinetics, validated methods to monitor compliance and, most importantly, led to the development of standardized preparations with defined concentrations of sulforaphane or its precursor, glucoraphanin (27–31). We sought to establish whether orally administered sulforaphane reaches the mammary gland and increases the functional capacity of antioxidative and detoxification enzymes in this tissue.

Materials and methods

Animals and sulforaphane treatments

Female Sprague–Dawley rats (10 weeks of age, 208–215 g) were purchased from Harlan (Indianapolis, IN) and maintained on an AIN-76A diet without...
ethoxyquin (Harlan Teklad, Madison, WI). Sprague–Dawley rats (10 weeks old) were used in this study, as the mammary glands at this age are quite mature, containing the highest known levels of the more differentiated alveolar buds (32,33). After 3 days of acclimatization, six rats were killed as controls whereas the remaining animals were gavaged with 150 μmol R,S-sulforaphane (LKT Laboratories, St Paul, MN) in 400 μl corn oil (Sigma–Aldrich, St Louis, MO). This dose of sulforaphane has been shown previously to inhibit the number, size and rate of development of mammary tumors in carcinogen-treated rats (16). At 0.5, 1, 2, 4, 8, 12, 24 and 48 h after treatment, three animals were killed (Figure 1A). From each rat, 4–5 ml of blood was obtained by cardiac puncture and transferred to a 15 ml conical polypropylene centrifuge tube containing disodium ethylenediaminetetraacetic acid for plasma sulforaphane determination. Mammary gland tissues were removed, immediately placed in liquid nitrogen and stored at –80°C for measurement of tissue sulforaphane levels and NQO1 enzymatic activity. Additional tissue was subsequently placed in RNA later (Ambion, Austin, TX) for gene expression analyses. Three additional animals were killed at 12, 24 and 48 h and mammary gland tissue was removed and placed in buffered 10% formalin for immunohistochemistry. All experiments were conducted in accordance with the standards established by the USA Animal Welfare Acts, set forth in National Institutes of Health guidelines and the Policy and Procedures Manual of the Johns Hopkins University Animal Care and Use Committee.

**Quantitative measurements of dithiocarbamates in urine and plasma** Measurement of sulforaphane distribution via dithiocarbamate (DTC) metabolites in biofluids was carried out using an established, well-validated cyclocondensation reaction (29).

**Pharmacokinetic analysis**

The measured concentrations of DTC metabolites in the rat plasma were fit into a non-compartmental extravascular model (NCA Model 200) using WinNonlin software (Pharsight Corporation, Mountain View, CA). The area under the curve (AUC), maximum concentration (Cmax), elimination rate constant (Kel) and elimination half-life (0.693/Kel) were obtained.

**Tissue DTC analysis**

The cyclocondensation reaction used to measure plasma DTCs was modified slightly for tissue analysis. Rat tissue (100 mg) was homogenized in 1 ml of 0.25 M sucrose and 10 mM Tris–HCl (pH 7.4) with a Duall grinder (Kontes Glass Co.) at 4°C for 2 min. Up to 0.6 ml of the crude homogenate was added directly to the cyclocondensation reaction mixtures and incubated as described previously (29). After incubation, the reaction mixture was centrifuged at low speed to remove insoluble materials. A 2 ml portion of the supernatant fraction was mixed with 2 ml water and loaded on a 50 mg Sep-Pak C18 Cartridge (Waters Corp., Milford, MA). The cartridge was then washed twice with 1 ml portions of 25% acetonitrile in water and subsequently eluted with 0.3 ml acetonitrile. The eluate was mixed with 0.3 ml water, and a 200 μl aliquot of the mixture was automatically injected (Waters Autosampler, Model 717 Plus) onto a reverse phase high performance liquid chromatography (HPLC) column (Partisil 10 ODS-2, 4.6 × 250 mm, Whatman, Clifton, NJ). The cyclocondensation product (1,3-benzodithiole-2-thione) was detected by a Waters Model 996 photodiode array detector at 365 nm. The DTC level of the tissue was adjusted to tissue weight.

**Gene expression analysis**

Total RNA was isolated using the RiboPure™ kit (Ambion, Austin, TX) and cDNA was synthesized using the iScript cDNA synthesis kit (Bio-Rad Laboratories, Hercules, CA). TaqMan polymerase chain reaction (PCR) assays were performed in triplicate on cDNA samples in 96-well optical plates on a Bio-Rad MyiQ Single-Color Real-Time PCR Detection System for either NQO1 or HO-1. For each 20 μl TaqMan reaction, 2 μl cDNA (10 ng) was mixed with 1 μl 20× TaqMan Gene Expression Assay Mix (Applied Biosystems, Foster City, CA), 10 μl 2 × iQ Supermix (Bio-Rad Laboratories) and 7 μl nuclease-free water (Promega Corporation, Madison, WI). Parallel assays for each sample were performed using glyceraldehyde-3-phosphate dehydrogenase (GAPDH) for normalization. The reaction was carried out using the following parameters: 50°C for 2 min, 95°C for 5 min and 40 cycles of 95°C for 15 s and 60°C for 1 min. Standard curves were prepared for each target using serial dilutions of a single sample. All TaqMan PCR data were captured using the MyiQ Optical System Software (Bio-Rad Laboratories). Relative quantification of gene expression changes in the rat was determined using the 2–DD method (34). Induction ratios are the ratios of treated to the control values. All time points >2 h (NQO1) or >1 h (HO-1) are significantly different from controls by t-tests (P < 0.05). Bars represent SEM (n = 3). (D) NQO1-specific enzymatic activity induction in the mammary gland (43,44) adjusted for protein concentration determined by the bicinchoninic acid assay (38). Activities from treated animals were normalized to controls to obtain an induction ratio. Time points >4 h are significantly different from controls as determined by t-tests (P < 0.05). Bars represent SEM (n = 3). (E) HO-1 immunostaining in the rat mammary gland after treatment with sulforaphane. Control (0 h) and treated (24 h). Original magnification, ×100. Scale bar = 50 μm.

Fig. 1. (A) Rat experiment design. Plasma and mammary glands were removed from three female Sprague–Dawley rats per time point following 150 μmol R,S-sulforaphane gavage at time 0. At select time points (*), mammary glands were removed from three additional animals for slide preparation. (B) Pharmacokinetics of sulforaphane in rat plasma and mammary gland. Shown are concentrations of DTCs (sulforaphane metabolites) versus time following a gavage of a single dose of 150 μmol R,S-sulforaphane in female 10-week-old Sprague–Dawley rats. The DTC level in the mammary gland was adjusted for tissue weight. Every time point between 0 and 48 h is significantly different from control animals by t-tests (P < 0.05). Bars represent SEM (n = 3). (C) Quantitative real-time PCR based measurements of rat mammary gland NQO1 and HO-1 transcripts. Relative quantification of gene expression changes was determined using the 2–DD method (34).
The homogenates were centrifuged at 14 000 r.p.m. for 20 min at 4°C. The NQO1 activity in the supernatant fractions was determined in a 250 μl assay in 96-well microtiter plates (35–37); 10, 20 and 40 μl of supernatants were added to the well and brought up to 50 μl with 0.25 M sucrose and 10 mM Tris–HCl (pH 7.4). In order to assess non-specific enzymatic activity, 50 μl of 0.3 mM dicumarol (NQO1 inhibitor) was added to parallel wells. To each well 200 μl assay solution containing 25 mM Tris–HCl pH 7.4, 0.5% bovine serum albumin, 0.025% Tween 20, 5 μM flavin adenine dinucleotide, 30 μM nicotinamide adenine dinucleotide phosphate, 1 μM glucose-6-phosphate, 2 U/ml glucose-6-phosphate dehydrogenase, 0.3 μg/ml 3-(4,5-dimethoxy-2-yl)-2,5-diphenyltetrazolium bromide and 25 mM menadione was added. The reaction rates were immediately measured at 610 nm for 5 min at room temperature by the microtiter plate reader. For each sample, the specific activity of NQO1 was derived by subtracting the initial reaction rates in wells containing no dicumarol from those in wells containing dicumarol. The protein level of the supernatant was measured by using the bicinchoninic acid assay (38). Activities from treated animals were normalized to controls to obtain an induction ratio.

**Immunohistochemical staining and scoring**

Staining for HO-1 was carried out using the DakoCytomation EnVision+ System (DakoCytomation, Carpinteria, CA). Briefly, deparaffinized slides were dehydrated and steamed, and endogenous peroxidase activity was quenched by incubation with Dako’s dual endogenous enzyme block. Slides were washed and incubated with Strexogen’s (Strexogen Bioreagents, Victoria, BC Canada) rabbit anti-human HO-1 polyclonal antibody (1:200 dilution) followed by a secondary anti-rabbit antibody-coated polymer peroxidase complex. Substrate/counter-stain (3,3’-diaminobenzidine) was applied and slides were counterstained with Gill-2 hematoxylin. Slides were scanned and six regions per slide were quantitatively scored using the ACIS II Automated Cellular Imaging System (Clariant, Aliso Viejo, CA) as described previously (39). Negative controls utilizing rabbit serum and secondary antibody alone revealed no immunostaining in any tissues examined.

**Human pilot study design**

Eight women with no prior history of cancer, recent antibiotic, aspirin, non-steroidal anti-inflammatory drugs (NSAID) or multivitamin use or with smoking history, who were scheduled to undergo elective reduction mammoplasty, were enrolled in a pilot clinical study. They had to complete a baseline dietary and risk factor questionnaire, be on a cruciferous-free diet for 3 days prior to surgery, complete a diet checklist of foods to avoid each day and collect their urine for the 12 h period prior to surgery. An hour before surgery, 10 ml of blood was drawn and the women were asked to drink 20 ml of a filtered broccoli sprout preparation containing sulforaphane. For this preparation, 3-day-old broccoli sprouts were grown by Sprouter Northwest (Kent, WA), extracted with hot water, treated with daikon and freeze dried under good manufacturing practice guidelines to produce a powder containing ~200 μmol of sulforaphane/g (Oregon Freeze Dry, Albany, OR). This powder was then stored in a freezer at –80°C. The amount of sulforaphane was confirmed prior to the preparation of each dose by a cyclocondensation assay (29). Blood and urine were also collected intra-operatively at the time when breast tissue was removed from the first breast. Breast tissue removed from both breasts was processed,

**Results**

**Sulforaphane metabolite distribution in the rat mammary gland**

In the rat mammary gland, DTC concentrations peaked at 18.8 pmol/mg tissue 1 h after oral administration of 150 μmol sulforaphane (Figure 1B). Similarly, DTC concentrations in the plasma peaked at 60 μM 1 h after dosing, declined rapidly and exhibited a minor second peak of 22 μM at 12 h (Figure 1B). The basis for this second peak is unclear but may relate to accumulation of glutathione conjugates (40,41). Pharmacokinetic parameters were generated using WinNonLin by fitting the plasma data to a non-compartmental extravascular input model (NCA Model 200) yielding an AUC of 491 h μmol/l, Cmax, of 60 μM, Kt of 0.1 h⁻¹ and an elimination half-life of 6.7 h. The peak plasma concentration was 3-fold higher than that reported by Hu et al. (42) following oral administration of 50 μmol sulforaphane to rats. These investigators also reported a shorter elimination half-life (2.2 h) perhaps reflecting measures of sulforaphane itself, rather than all of its biotransformation products (DTCs).

**Induction of cytoprotective gene transcripts in the rat mammary gland**

Quantitative real-time PCR assays were used to measure levels of NQO1 and HO-1 transcripts to determine the extent of the pharmacodynamic action of sulforaphane in the rat mammary gland. A maximal 12-fold induction of NQO1 transcripts was observed in the mammary gland 12 h after dosing and there was significant induction as early as 2 h (P = 0.0001) (Figure 1C). A biphasic pattern of HO-1 transcript induction was observed, with an initial peak at 2 h followed by a subsequent peak at 12 h (Figure 1C). The minimal time to statistically significant HO-1 induction was 1 h (P = 0.0001). Transcript levels of GAPDH did not significantly change in any of the tissues and therefore served as an appropriate control.

**Induction of NQO1 enzymatic activity in the rat mammary gland**

A microtiter plate assay was used to determine whether a single dose of sulforaphane leads to measurable induction of NQO1 enzymatic activity in the rat mammary gland (43,44). A maximal 2.8-fold induction of NQO1 enzymatic activity in the mammary gland was observed 24 h (P = 0.0001) after dosing with minimal time to statistically significant induction at 4 h (P = 0.0001) (Figure 1D).

**Localization of increased HO-1 immunostaining to rat mammary epithelial cells**

Immunostaining assays for HO-1 were used to localize the induction of cytoprotective enzymes. Minimal HO-1 immunostaining was noted throughout the control samples. Strong immunostaining for HO-1 was observed in the mammary gland 24 h after a single sulforaphane dose with considerable staining evident in mammary ductal and glandular epithelial cells (Figure 1E). Using the ACIS II Automated Cellular Imaging System the slides were scanned and six regions per slide were scored with 4-fold increased HO-1 staining confirmed in the treated mammary glands compared with controls (mean (SD) ACIS percentage × intensity score, 8640 (3662) versus 2168 (1121), P = 0.002).

**Sulforaphane metabolite distribution in human breast tissue**

To determine whether sulforaphane is bioavailable in human breast tissues, eight women undergoing reduction mammoplasty were enrolled in a proof of principle pilot study (Figure 2). All eight women were African American. Their average age was 30 years (21–50) and seven were pre-menopausal. Their mean pre-operative weight was 99 kg (68.3–158.8). They all consumed a broccoli sprout preparation containing 200 μmol of sulforaphane on average 50 min (27–75) prior to the start of surgery. As expected, the broccoli sprout preparation was well tolerated. This dose and schedule were chosen because previous work demonstrated a peak plasma concentration of 2.0 μM DTCs at 1 h after ingestion (29). The average time between the dose and removal of the right and left breast tissue was similar, 93 ± 17 and 103 ± 33 min, respectively. The mean epithelial-stromal-enriched breast tissue DTC concentration was 1.45 ± 1.12 and 2.00 ± 1.95 μM/ml of tissue for the right and the left breast, respectively (Table I). The mean pre-dose urinary DTC concentration was 4.07 ± 5.22 μM with a level of 158.85 ± 93.89 μM obtained at the time of tissue removal, consistent with isothiocyanate ingestion. The mean pre-dose plasma DTC concentration was negligible (0.01 ± 0.02 μM), with a post-dose level of 0.92 ± 0.72 μM.
Measurement of cytoprotective gene transcripts and NQO1 enzymatic activity in human breast tissue

It was not possible to evaluate pharmacodynamic effect in breast tissue because of the short interval between ingestion of the broccoli sprouts preparation and breast resection. Nonetheless, it was possible to detect NQO1 and HO-1 transcripts and NQO1 enzymatic activity in the mammary tissue of all subjects. For NQO1, a mean ΔCt (NQO1–GAPDH) of 4.50 ± 0.70 in the right breast and 4.23 ± 0.63 in the left breast was determined. For HO-1, a mean ΔCt (HO-1–GAPDH) of 6.30 ± 0.41 in the right breast and 6.26 ± 0.34 in the left breast was determined. The transcript levels were highly correlated (correlation coefficient = 0.78) between breasts for both NQO1 and HO-1. Mean NQO1 enzymatic activity was determined with levels of 0.21 ± 0.1 mOD/µg/min in the right breast and 0.20 ± 0.1 mOD/µg/min in the left breast.

Discussion

This study was carried out to investigate the pharmacokinetics and pharmacodynamics of sulforaphane in the rat mammary gland with hopes of identifying biomarkers that can be used to assess the chemopreventive efficacy of broccoli sprouts in human clinical trials. For the first time, we measured sulforaphane metabolites in mammary tissue from 10-week-old female Sprague–Dawley rats as early as 30 min following a single oral 150 µmol dose of sulforaphane. This was followed by significant induction of NQO1 and HO-1 cytoprotective gene transcripts in the rat mammary gland. This response is consistent with prior studies examining NRF2-mediated transcripts induced by sulforaphane in the rodent small intestine and liver (42,45). The pattern and timing of induction varied between transcripts. The more rapid and biphasic induction of HO-1 transcripts observed in sulforaphane-induced rat mammary cells (42) is consistent with its mechanism of action. While induction of NQO1 would have been desirable, published data and pilot studies showed substantial NQO1 staining in untreated rat and human mammary ductal epithelial cells (51). Together, our studies demonstrate that sulforaphane distributes to the breast epithelial cells in vivo and exerts a pharmacodynamic action in these target cells consistent with its mechanism of chemoprotective efficacy. Such efficacy, coupled with earlier randomized clinical trials demonstrating the safety of repeated doses of broccoli sprout preparation containing 200 µmol of sulforaphane (27–31), supports further evaluation of broccoli sprouts in the chemoprevention of breast and other cancers.

Table I. Human sulforaphane pharmacokinetics

<table>
<thead>
<tr>
<th>Specimens (units)</th>
<th>Pre-dose</th>
<th>Post-dose</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breast tissue</td>
<td>—</td>
<td>Left: 2.00 ± 1.95; Right: 1.45 ± 1.12</td>
</tr>
<tr>
<td>(µmol/mg tissue)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Urine (µM)</td>
<td>4.07 ± 5.22</td>
<td>158.85 ± 93.89</td>
</tr>
<tr>
<td>Plasma (µM)</td>
<td>0.01 ± 0.02</td>
<td>0.92 ± 0.72</td>
</tr>
</tbody>
</table>

Data, mean (n = 8) ± SE.

In humans, >85% of invasive breast cancers originate in ductal epithelial cells (50). Using immunostaining assays, we were able to detect significant HO-1 induction in rat mammary ductal and glandular epithelial cells 24 h after dosing. HO-1 was chosen as several studies revealed little to no immunostaining in both normal human breast tissue (http://www.proteinatlas.org) and established breast epithelial cell lines (51). While immunostaining of NQO1 would have been desirable, published data and pilot studies showed substantial NQO1 staining in untreated rat and human mammary ductal epithelial cells (52) minimizing the capacity to detect induction. In a proof of principle clinical study, we were also able to demonstrate the presence of DTCs in human breast tissue after a single dose of a broccoli sprout preparation containing 200 µmol of sulforaphane. In addition, we have been able to measure NQO1 and HO-1 transcripts, as well as NQO1 enzymatic activity in human breast tissue. Together, our studies demonstrate that sulforaphane distributes to the breast epithelial cells in vivo and exerts a pharmacodynamic action in these target cells consistent with its mechanism of chemoprotective efficacy. Such efficacy, coupled with earlier randomized clinical trials demonstrating the safety of repeated doses of broccoli sprout preparations (27–31), supports further evaluation of broccoli sprouts in the chemoprevention of breast and other cancers.

Acknowledgements

This research was supported by National Institutes of Health grants, Breast Spore P50 CA088843, Breast Cancer Research Foundation and the Lewis B. and Dorothy Cullman Foundation. K.V. is a recipient of an ASCO Career Development Award and a KO7 Preventive Oncology Academic Award CA11948 from the National Cancer Institute. B.S.C. was supported by T32 ES07141. We thank Patrick Dolan for assistance with animal care and Brian Schofield, Jessica Hicks, Kristen Lecksell and Angelo DeMarzo for their assistance with immunohistochemical assays. We thank Katherine Stephenson and Kristina L.Wade for assistance in making the broccoli preparation and Deborah Santor, Anita Mayfield and Amy Gross for assistance with the clinical studies.
Conflict of Interest Statement: None declared.

References


Received November 30, 2006; revised February 4, 2007; accepted February 22, 2007